



0040-4020(94)00530-3

## Limonoids from *Carapa grandiflora* (Meliaceae)

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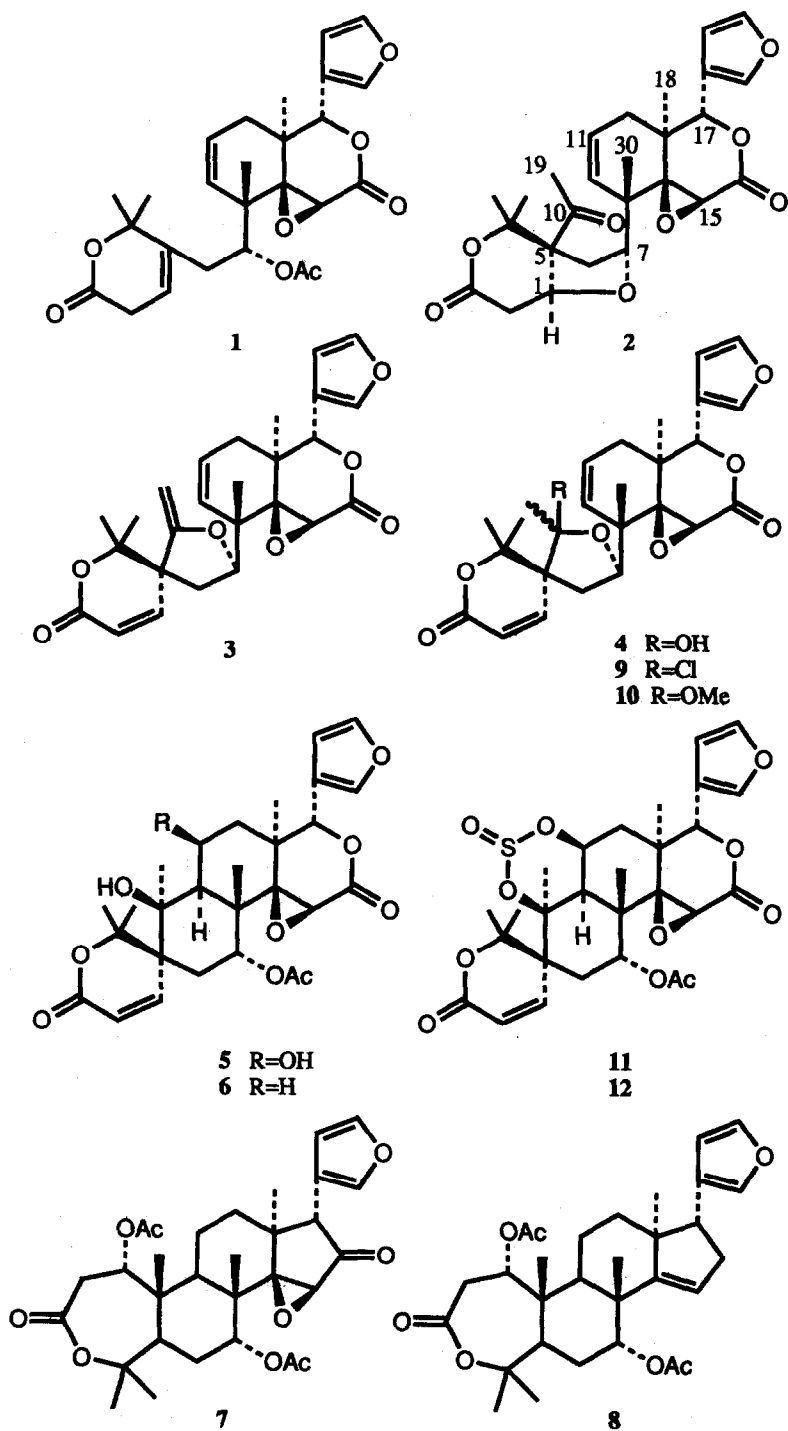
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**Abstract:** Eight limonoids, carapolides C 1, D 3, E 4, F 5, G 6, H 7 and I 8 and evodoulone 13 have been isolated from the extracts of the seeds of *Carapa grandiflora* (Meliaceae). Compounds 2, 3, and 4 are new C-9, C-10 cleaved tetranortriterpenoids while 5 and 6 are novel tetranortriterpenoid spiro lactones. Finally, carapolides H 7 and I 8 are new ring A cleaved derivatives with an intact ring D. All their structures were determined from spectroscopic data and chemical correlations.

The seeds of *Carapa grandiflora* sprague (Meliaceae), popularly known in Cameroon as "engang", are used in traditional medicine as a febrifuge and for the treatment of arthritis, general fatigue and skin diseases<sup>1</sup>. As part of our continuing search for plant filaricides related to carapolide A 1, which had shown strong *in vitro* activity against *Onchocerca volvulus* microfilariae<sup>2,3</sup>, we have investigated the extract of the seeds of *C. grandiflora* and obtained eight tetranortriterpenoids, some of which are related to carapolide A 1, but not compound 1 itself. A preliminary communication<sup>4</sup> reported the structural elucidation of three of them, carapolide D 3, E 4, and F 5. The present paper reports the isolation and structure determination of four additional limonoids from the seeds and presents full experimental details concerning 3-5 and 13.

An acetone extract of the defatted seeds was subjected to repeated column chromatography, affording compounds 2, 3, 4, 5, 6, 7, 8, and 13 in yields of 0.002, 0.001, 0.23, 0.01, 0.02, 0.018, 0.002, and 0.3%, respectively. Evodoulone 13, previously obtained from *C. procera*<sup>5</sup>, was characterised by direct comparison (mmp,  $[\alpha]_D$ , IR and NMR) with an authentic sample.

The structures of carapolides C 2<sup>6</sup>, D 3, and E 4 have already been reported<sup>4</sup>. They constitute further members of the rare group of tetranortriterpenoids with a C-9, C-10 cleaved carbon skeleton. Attempts were made to correlate chemically these three compounds. Thus treatment of carapolide E 4 with p-toluene sulfonic acid yielded carapolide C 2, identical in all respects with the natural sample. Efforts to convert carapolide E 4 into carapolide D 3 through dehydration with thionyl chloride in dry pyridine at 0°C were, however, unsuccessful and led to the formation of the chloride 9. The IR spectrum of 9 lacked a hydroxyl absorption band and, though its <sup>1</sup>H NMR spectrum was very similar to that of 4, the hydroxyl proton resonance at δ 2.41 ppm in the spectrum of 4 was absent. As expected, reaction of carapolide E 4 with acidic methanol yielded the methyl acetal 10. The <sup>1</sup>H NMR spectral data of the acetal (experimental section) were consistent with structure 10. Paucity of material precluded attempts to transform 3 into 4 by hydration.



For carapolide F **5**, EI-LRMS and elemental analysis established the molecular formula  $C_{28}H_{34}O_{10}$ . Its spectroscopic properties clearly indicate that it has an intact ring B and are consistent with the spirostructure **5**. Thus it has in addition to a  $\beta$ -substituted furan and a ring D epoxy lactone, resonances ( $^1H$  and  $^{13}C$  NMR spectra Tables 1 and 3) for five tertiary methyl groups, a ring A  $\alpha\beta$ -unsaturated  $\delta$ -lactone and a secondary acetate ( $\delta_H$  2.01 (3H, s), 4.55 (dd, J 2.3, 3.7 Hz, H-7);  $\delta_C$  170.2 (s) and 74.3 (d, C-7)). H-7 is coupled to an isolated methylene group 2H-6 ( $\delta_H$  2.63 (dd, J 15.2 and 2.3 Hz) and 2.13 (dd, J 15.2 and 3.7 Hz)) showing that C-5 is fully substituted. There are two additional oxygenated carbons,  $\delta_C$  78.6 (s, C-10) and  $\delta_C$  65.4 (d, C-11), both bearing hydroxyl groups. H-11 appears as a multiplet ( $\delta_H$  4.85) and is coupled to H-9 ( $\delta_H$  2.50 (d, J 5.0 Hz)) and to the C-12 methylene group ( $\delta_H$  1.79 (dd, J 13.5 and 9.4 Hz, H-12 $\alpha$ ) and 2.21 (dd, J 13.5 and 7.4 Hz, H-12 $\beta$ )). The coupling data suggest that the 11-hydroxyl group is pseudoequatorial ( $\beta$ ) with ring C adopting a boat conformation and the C-7 acetoxyl group is axial in a chair ring B. The relative stereochemistry of the methyl group at C-10 was defined as equatorial by strong nOe interactions between Me-19 and H-1 (4.6%), Me-19 and H-11 (7.3%), and Me-19 and H-9 (1.8%). All other nOe enhancements, particularly those observed between H-9 and H-1 (9.6%), H-9 and H-11 (5.2%) and H-9 and Me-18 (9.4%) are fully consistent with structure **5**.

Treatment of carapolide F **5** with  $SOCl_2$  in pyridine afforded two diastereoisomeric cyclic sulphites **11** and **12**, differing in the configuration at sulphur, which were separated by preparative TLC. The  $^1H$  NMR spectra of these derivatives (Experimental) are similar in chemical shifts and coupling constants to that of **5**, apart from showing downfield shifts of H-11 and the C-10 methyl. The major differences between the spectra of the sulphites **11** and **12** involve the shifts of H-11 ( $\delta_H$  5.49 and 5.78), H-9 (2.46 and 2.59), and the C-10 methyl group ( $\delta_H$  1.99 and 2.20). The formation of these cyclic esters supports the assigned structure **5** for carapolide F.

Carapolide G **6**,  $C_{28}H_{34}O_9$ , by elemental analysis, has IR bands at 3500, 1760, 1740, 1715, and 1700  $cm^{-1}$ , indicative of hydroxyl and carbonyl functionality. All the spectroscopic data are appropriate for an intact ring B skeleton and are consistent with the spiro lactone structure **6**. The  $^1H$  NMR spectrum of **6** is almost superimposable on that of carapolide F **5**, except for the appearance in the up-field region of the spectrum of a multiplet of four protons at  $\delta$  1.50-1.95 and the absence of the oxymethine proton signal at  $\delta$  4.80. These data suggest that the C-11 hydroxyl of **5** is absent in carapolide G **6**. This was confirmed by the  $^{13}C$  NMR spectrum of **6**. Thus the methine resonance at  $\delta$  65.7 (C-11) in the  $^{13}C$  spectrum of **5** is replaced by a methylene signal at  $\delta$  14.5 in that of carapolide G **6**. The  $^1H$  and  $^{13}C$  NMR assignments, made using  $^1H$ - $^1H$  COSY and  $^1H$ - $^{13}C$  correlations, are fully consistent with structure **6**. The  $\alpha$  (equatorial) orientation of the C-10 methyl group (Me-19) was established by nOe experiments, key nOe interactions being observed between Me-19 and H-1 and Me-19 and H-9. All other nOe enhancements, including those between Me-18 and H-9, H-9 and H-1, and H-9 and Me-18 support the relative stereochemistry as in **6**. The stereochemistry of carapolide F **5** and G **6** at C-10 is identical with that of pedonin **14** whose structure was established by X-ray crystallography<sup>7</sup>.

The tetranortriterpenoid nature of carapolide H **7** and its close relationship to  $7\alpha$ -acetoxyl dihydronomilin **15**<sup>8</sup> is apparent from its spectroscopic data. It analysed for  $C_{30}H_{38}O_9$ , and showed three strong carbonyl absorptions,  $\nu_{max}$  1740, 1720, and 1700  $cm^{-1}$  in its IR spectrum. Its  $^1H$  NMR spectrum (Table 4) revealed resonances for five tertiary methyl groups, two acetoxy groups, and a secondary epoxide proton ( $\delta_H$  3.39, H-15). In addition to the characteristic  $\beta$ -substituted furan signals, two oxymethine resonances are clearly discernable at  $\delta$  4.95 (d, J 7.0 Hz, H-1) and 4.65 (bt, J 3.0 Hz, H-7), respectively. A proton resonating as

Table 1. 50.32 MHz  $^{13}\text{C}$  NMR Data for Compounds 2-5.

Carbon #	2 <sup>a</sup>	3 <sup>a</sup>	4 <sup>a</sup>	5 <sup>b</sup>
1	75.8d	148.7d	149.7d	154.4d
2	35.7t	118.3d	120.5d	117.0d
3	169.7s	163.1s	163.3s	165.2s
4	81.7s	84.0s	85.1s	90.8s
5	66.8s	52.1s	55.4s	47.4s
6	33.2t	35.7t	36.6t	31.3t
7	81.5d	81.2d	80.3d	74.4d
8	42.0s	41.6s	42.2s	41.0s
9	131.3d	131.0d	131.7d	39.9s
10	206.3s	159.8s	105.2s	78.6s
11	122.7d	123.0d	122.4d	65.4d
12	32.2t	32.1t	32.1t	38.2t
13	38.1s	38.1s	38.1s	38.4s
14	65.2s	66.7s	67.7s	70.1s
15	52.5d	52.5d	52.7d	57.1d
16	167.0s	166.9s	167.3s	168.8s
17	77.8d	77.8d	77.8d	78.8d
20	120.1s	120.0s	120.3s	120.6s
21	141.0d	141.0d	141.0d	141.8d
22	109.9d	109.9d	109.9d	110.2d
23	143.2d	143.2d	143.2d	143.7d
28	27.0 <sup>*</sup> q	25.0 <sup>*</sup> q	26.7 <sup>*</sup> q	27.6q
29	26.5 <sup>*</sup> q	23.3 <sup>*</sup> q	24.9 <sup>*</sup> q	27.6q
30	16.7q	16.8q	18.2q	19.8
18	16.1q	16.1q	16.1q	16.3
19	28.6q	86.3t	24.9q	26.1q
OAc	-	-	-	170.2s
	-	-	-	21.1q
OAc	-	-	-	-

\* Assignments are interchangeable  
<sup>a</sup>  $\text{CDCl}_3$ , <sup>b</sup>  $\text{CDCl}_3 + \text{CD}_3\text{OD}$

Table 2. 50.32 MHz  $^{13}\text{C}$  NMR Data for Compounds 6-8.

Carbon #	6 <sup>a</sup>	7 <sup>a</sup>	8 <sup>b</sup>	8 <sup>c</sup>
1	153.7d	72.0d	70.9d	71d
2	116.4d	35.6t	34.9t	34.9t
3	164.1s	172.6s	170.3s	170.2s
4	89.7s	87.2s	85.5s	85.5s
5	48.1s	45.4d	44.1d	44.1d
6	30.4t	27.5t	26.3t	26.4t
7	72.8d	74.8d	74.2d	74.3d
8	40.8s	43.5s	41.8s	41.8s
9	37.4d	37.9d	36.0d	36.0d
10	76.0s	43.4s	44.2s	44.3s
11	14.5t	16.7t	16.3t	16.3t
12	25.1t	30.2t	32.8t	32.9t
13	38.5s	45.3s	47.0s	47.1s
14	67.7s	73.2s	158.6s	158.1s
15	56.8d	57.7d	119.1d	119.1d
16	168.0s	209.9s	34.3t	34.4t
17	78.1d	51.8d	51.2d	51.3d
20	120.0s	117.6s	124.4s	124.5s
21	140.9d	142.4d	139.6d	139.6d
22	109.8d	111.8d	110.9d	111.0d
23	143.0d	143.4d	142.6d	142.6d
C-Me	27.1q	34.7q	34.4q	34.4q
C-Me	26.7q	25.4q	27.2q	27.2q
C-Me	25.3q	23.4q	23.6q	23.6q
C-Me	17.5q	19.7q	19.8q	19.8q
C-Me	16.4q	16.3q	15.2q	15.2q
OAc	169.5s	171.1s	170.0s	170.0s
	20.9q	21.5 <sup>*</sup> q	21.1 <sup>*</sup> q	21.2 <sup>*</sup> q
OAc	-	170.7s	169.8s	169.7s
	-	21.0 <sup>*</sup> q	20.7 <sup>*</sup> q	20.7 <sup>*</sup> q

<sup>a</sup>  $\text{CDCl}_3 + \text{CD}_3\text{OD}$ , <sup>b</sup>  $\text{CDCl}_3$ , <sup>c</sup> Values from ref. 10; <sup>\*</sup> Assignments are interchangeable

Table 3. <sup>1</sup>H NMR Data for Compounds 2-5

Carbons	2 <sup>a</sup>	3 <sup>a</sup>	4 <sup>a</sup>	5 <sup>b</sup>
1	4.83 (dd, 7.3, 2.2)	6.58 (d, 9.7)	6.78 (d, 10.0)	6.78 (d, 10.0)
2	3.03 (dd, 18.3, 7.3) 2.68 (dd, 18.3, 2.2)	5.98 (d, 9.7)	6.11 (d, 10.0)	5.91 (d, 10.0)
6	2.25 (d, 8.2)	1.85 (d, 8.0)	1.80 (d, 8.0)	2.13 (dd, 15.2, 3.7) 2.63 (dd, 15.2, 2.3)
7	4.17 (t, 8.2)	4.34 (dd, 7.6, 8.7)	4.13 (dd, 8.0, 8.7)	4.55 (dd, 2.2, 3.7)
9	5.52 (m)	5.84 (dd, 10.3, 2.5)	5.85 (m)	2.15 (d, 5.0)
11	5.57 (m)	5.76 (m)	5.77 (m)	4.85 (m)
12	1.80 (dd, 18.1, 2.2) 2.15 (d, 18.1)	-	-	1.79 (dd, 13.5, 9.4) 2.21 (dd, 13.5, 7.4)
15	3.72 (s)	3.88 (s)	3.77 (s)	3.50 (s)
17	5.57 (s)	5.59 (s)	5.60 (s)	5.62 (s)
19	2.30 (s)	4.17 (d, 2.0) 4.49 (d, 2.0)	-	-
21	7.40 (m)	7.40 (m)	7.40 (m)	7.43 (m)
22	6.35 (m)	6.35 (m)	6.35 (m)	6.35 (m)
23	7.40 (m)	7.40 (m)	7.40 (m)	7.40 (m)
OAc	-			2.01
OH	-		2.41 (s)	
C-Me	0.85		0.81	1.16
C-Me	1.00	0.88	1.08	1.55
C-Me	1.40	1.11	1.50	1.59
C-Me	1.55	1.48	1.52	1.64
C-Me		1.50	1.63	1.65

<sup>a</sup>500 MHz; <sup>a</sup> CDCl<sub>3</sub>; <sup>b</sup> CDCl<sub>3</sub> + CD<sub>3</sub>OD

Multiplicities and values of coupling constants (Hz) are given in parentheses.

Table 4. <sup>1</sup>H NMR Data for Compounds 6-8.

Carbons	6 <sup>b</sup>	7 <sup>b</sup>	8 <sup>a</sup>	8 <sup>c</sup>
1	6.85 (d, 10.0)	4.91 (dd, 6.5,1.5)	4.80 (brt)	4.82 (d, 7.0)
2	5.80 (d, 10.0)	3.05 (d, 7.0) 2.95 (dd, 7.0, 2.0)	3.10 (m)	3.12 (d, 15.0) 3.17 (dd, 15.0, 7.0)
5		2.46(dd,13.2,2.6)	2.50 (m)	2.52 (m)
6	2.51 (dd, 15.0, 4.0) 2.05 (dd, 15.0, 3.0)	1.80-2.15 (m)	1.94 (m)	1.95 (m)
7	4.45 (t, 3.0)	4.65 (t, 3.0)	5.17 (t, 3.0)	5.19 (brs)
9	2.46 (t,9.5)	2.92 (dd, 11.7,3.2)	2.53 (m)	2.56 (m)
11	1.50-1.90 (m)	1.90-2.15 (m)	1.40-1.60 (m)	1.42 (m) 1.57 (m)
12	1.50-1.90 (m)	1.55-1.75 (m)	1.40-1.60 (m)	1.57 (m)
15	3.38 (s)	3.39 (s)	5.34 (dd, 3.0, 2.0)	5.36 (brs)
17	5.52 (bs)	3.86 (s)	2.77 (t, 7.8)	2.73 (t, 7.5)
21	7.32 (bs)	7.50 (t, 2.0)	7.20 (m)	7.23 (m)
22	6.23 (bs)	6.22 (dd, 2.0, 1.0)	6.20 (dd, 2.0, 1.0)	6.26 (s)
23	7.32 (m)	7.40 (m)	7.35 (t, 2.0)	7.37 (brs)
OAc	1.93 (s)	2.03 (s)	1.98 (s)	2.01(s)
OAc		2.05 (s)	2.06 (s)	2.11 (s)
C-Me	1.10 (s)	0.98 (s)	1.15 (s)	1.17 (s)
C-Me	1.23 (s)	1.14 (s)	1.19 (s)	1.21(s)
C-Me	1.26 (s)	1.16 (s)	1.38 (s)	1.40 (s)
C-Me	1.50 (s)	1.36 (s)	1.49 (s)	1.51 (s)
C-Me	1.55 (s)	1.47 (s)	0.72 (s)	0.75 (s)

<sup>a</sup> CDCl<sub>3</sub>; <sup>b</sup> CDCl<sub>3</sub> + CD<sub>3</sub>OD

Multiplicities and values of coupling constants (Hz) are given in parentheses

<sup>c</sup> values from ref. 10

a singlet at  $\delta_{\text{H}}$  3.86 was assigned to H-17, in view of its nOe to the furan H-22. The up-field nature of the chemical shift of this proton in **7** relative to that ( $\delta_{\text{H}}$  5.60) observed for the same proton in 7 $\alpha$ -acetoxy dihydronomilin<sup>8</sup> suggested that carapolide H **7** has an intact ring D. Structure **7** was thus attributed to carapolide H. Its <sup>13</sup>C and <sup>1</sup>H NMR spectral data were assigned using <sup>1</sup>H-<sup>1</sup>H COSY and <sup>1</sup>H-<sup>13</sup>C correlation experiments (Tables 2 and 4). NOe experiments support the relative stereochemistry of **7** as shown. In particular, nOes from Me-19 to H-1 $\beta$  (3.0%) and from Me-30 to H-7 $\beta$  (6.0%) establish the configurations of the acetate groups.

Carapolide I **8**, C<sub>30</sub>H<sub>40</sub>O<sub>7</sub>,  $\nu_{\text{max}}$  1730, 1725, and 875 cm<sup>-1</sup>, has the same carbon skeleton as **7** but lacks the 14, 15-epoxide and the 16-keto group. All its spectroscopic data including <sup>1</sup>H and <sup>13</sup>C NMR (Tables 2 and 4) are consistent with structure **8**. {Compound **8** was presented at the 4th NAPRECA International Symposium on Natural Product Development held in Addis Ababa, Ethiopia, in September 1991<sup>9</sup>. However, at the time of submission of this manuscript, we came across a reference in which the same compound has been communicated from a different source as kihadalactone A<sup>10</sup>}.

A possible biogenetic pathway leading to the formation of the carapolides of *Carapa grandiflora* can now be proposed (scheme 1). The spirolactone carapolide G **6** appears to be the biogenetic progenitor. Hydroxylation of **6** yields carapolide F **5**. A retro-Prins reaction of **5** results in cleavage of the C-9, C-10, bond with formation of the ring C double bond and a methyl ketone as in **16**. Subsequent hydrolysis of the C-7 acetoxy group followed by internal hemiacetal formation involving the methyl ketone and the C-7 hydroxyl group then leads directly to carapolide E **4** and indirectly to D **3** by dehydration. Carapolide C **2** can arise from the 7-deacetyl derivative of **16**, by addition of the C-7 hydroxyl group to the ring A unsaturated lactone to form the cyclic ether.

## EXPERIMENTAL

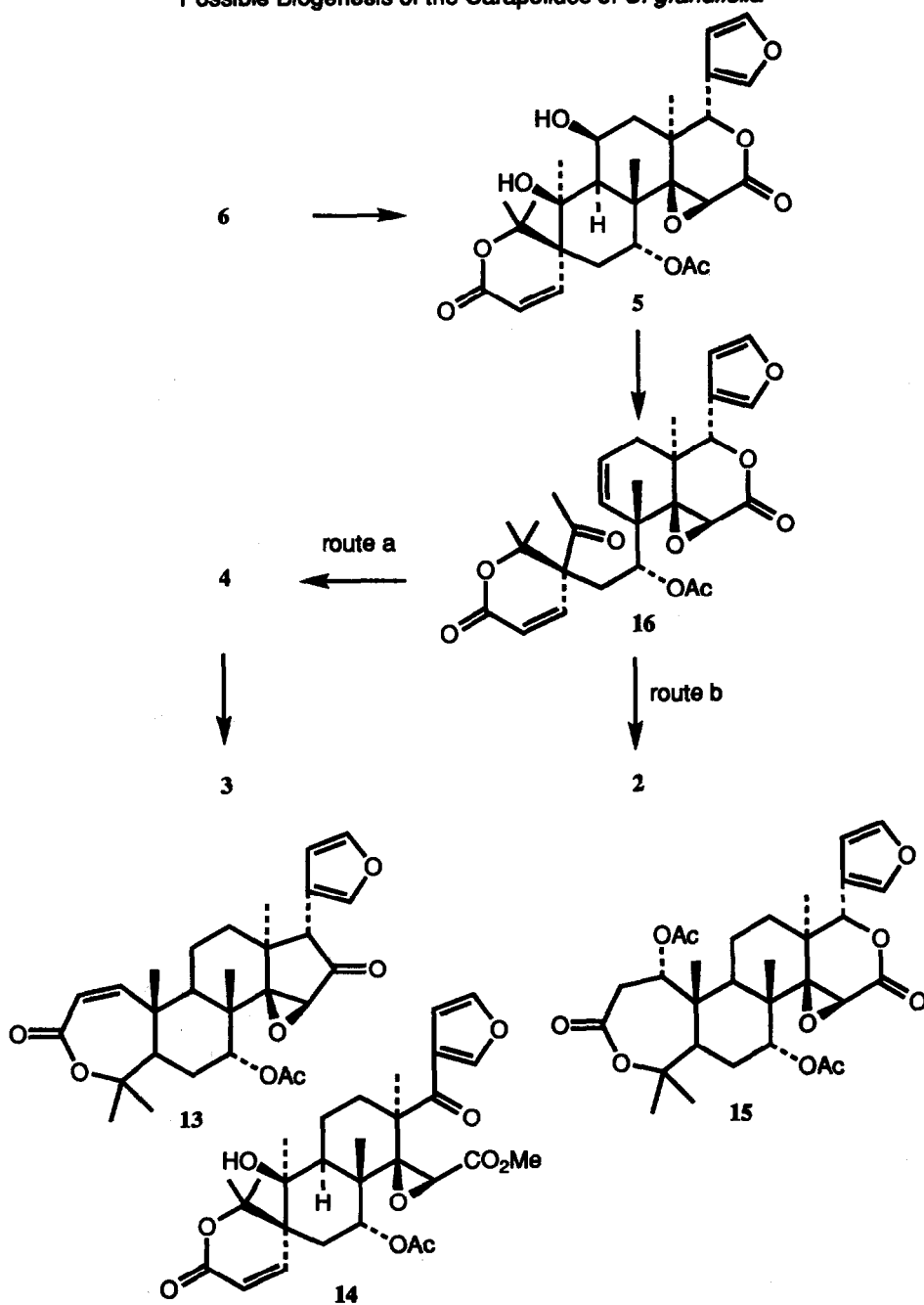
Melting points were measured on a Kofler hot stage apparatus and are uncorrected. IR spectra were recorded on a Perkin-Elmer 727B spectrophotometer. NMR spectra in CDCl<sub>3</sub> solutions were run at 25°C in pulsed Fourier transform mode either on a Varian XL-100 spectrometer (25.16 MHz for <sup>13</sup>C, shifts relative to Me<sub>4</sub>Si at  $\delta$  0.00) or on a Bruker WP 200SY spectrometer (200.13 MHz for <sup>1</sup>H, shifts relative to CHCl<sub>3</sub> at  $\delta$  7.25; 50.32 MHz for <sup>13</sup>C, shifts relative to CDCl<sub>3</sub> at  $\delta$  77.00). Mass spectra were determined using an MS 902S instrument. Kieselgel 60 (0.063-0.200 mm, Merck) was used for column chromatography, and (0.0400-0.063 mm, Merck) for Medium Pressure liquid chromatography (Baekström Separo AB Apparatus). TLC was run on Merck GF<sub>254</sub> silica gel pre-coated plates (0.2 mm thickness).

### Isolation of Limonoids from *Carapa grandiflora*

The powdered sun-dried seeds (3 kg), obtained from the fruits of *C. grandiflora* collected in August 1983 from Yaoundé, Cameroon, were extracted successively with *n*-hexane (10 l) and acetone (10 l) in a Soxhlet apparatus. The hexane extract was concentrated *in vacuo* to leave an oil (1100 g) while evaporation of the solvent under reduced pressure from the acetone extract yielded a brown gum (330 g). A sample of the acetone extract (110 g) was dissolved in CHCl<sub>3</sub> (60 ml) and chromatographed on a column of Si gel (1.5 kg). Elution started with *n*-hexane and continued stepwise through *n*-hexane/EtOAc mixtures, EtOAc, and EtOAc/MeOH mixtures. The eluate was collected in 250 ml fractions which were combined using TLC comparisons in appropriate solvent systems. From this chromatographic separation evodoulone<sup>5</sup> **13** (1.8 g),



Scheme 1  
Possible Biogenesis of the Carapolides of *C. grandifolia*



carapolide D 3 (120 mg), carapolide I 8 (600 mg), carapolide E 4 (1.5 g), a mixture of 2.5 g of carapolides - C 3, E 4, and G 6, and a mixture of 2.7 g of carapolides F 5 and H 7 were obtained. Further purification of the various mixtures by MPLC using the recently developed Baeckström AB Separo Columns (15 and 25 mm diameter) with a continuous gradient hexane-EtOAc-MeOH afforded pure carapolides C 2 (85 mg), F 5 (300 mg), G 6 (480 mg), and H 7 (380 mg). Evodoulone 13 was identified by direct comparison with an authentic specimen previously available in our laboratory from *Carapa procera* while the new limonoids were characterised as follows.

**Carapolide C 2.** Colourless glass;  $[\alpha]_D^{25} + 102^\circ$  (c, in  $\text{CHCl}_3$ ); IR (KBr)  $\nu_{\text{max}}$  1752, 1710, 1600, 878  $\text{cm}^{-1}$ ; EI-HRMS  $m/z$  470.1948,  $\text{C}_{26}\text{H}_{30}\text{O}_8$  requires  $m/z$  470.1931.  $^1\text{H}$  and  $^{13}\text{C}$  NMR see Tables 1 and 3.

**Carapolide D 3.** Prisms from EtOAc/hexane, mp 234-236 $^\circ$ ;  $[\alpha]_D^{20} - 362^\circ$  (c = 1.8, acetone); EI-HRMS:  $m/z$  452.1844,  $\text{C}_{26}\text{H}_{28}\text{O}_7$  requires 452.1833; IR (KBr)  $\nu_{\text{max}}$  3150, 3070, 1750, 1710, 1650, 1630, 1462, 1400, 1380, 1300, 1260, 1220, 1160, 1120, 1100, 1060, 1025, 990, 940, 905, 880, 810  $\text{cm}^{-1}$ .  $^1\text{H}$  and  $^{13}\text{C}$  NMR see Tables 1 and 3.

**Carapolide E 4.** Mp 178-180 $^\circ$  (ex EtOAc/hexane);  $[\alpha]_D^{20} + 106^\circ$  (c, 4.3 in acetone): IR (KBr)  $\nu_{\text{max}}$  3370, 3140, 3080, 1735, 1720, 1450, 1380, 1280, 1165, 1135, 1080, 1040, 980, 878, 825, 810, 740  $\text{cm}^{-1}$ . EI-HRMS:  $m/z$  470.1950,  $\text{C}_{26}\text{H}_{30}\text{O}_8$  requires  $m/z$  470.1939. Found: C, 66.29; H, 6.52.  $\text{C}_{26}\text{H}_{30}\text{O}_8$  requires: C, 66.37; H, 6.43.  $^1\text{H}$  and  $^{13}\text{C}$  NMR see Tables 1 and 3.

#### Conversion of Carapolide E 4 to Carapolide C 2

Carapolide E 4 (150 mg) and *p*-toluene sulfonic acid (500 mg) were dissolved in dry benzene and the solution refluxed in a Dean and Stark apparatus for 15 min. The reaction mixture was then cooled, washed with aqueous  $\text{Na}_2\text{CO}_3$  solution (5%), dried with anhydrous  $\text{Na}_2\text{SO}_4$  and evaporated *in vacuo*. Purification by MPLC gave carapolide C 2 (132 mg) identical in all respects with the natural sample.

#### Attempted conversion of Carapolide E 4 into Carapolide D 3

**Carapolide E chloride 9.** To a solution of carapolide E 4 (30 mg) in dry pyridine (2 ml) at 0 $^\circ\text{C}$  was added  $\text{SOCl}_2$  (8 drops). After 2h, the reaction mixture was warmed up to r.t., diluted with water (50 ml) and extracted with  $\text{CHCl}_3$ . Preparative TLC of the product afforded the chloride 9 (25 mg) as a colourless glass; IR (KBr)  $\nu_{\text{max}}$  3140, 1750, 1730, 880  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR (200 MHz,  $\text{CDCl}_3$ ) 6.8 (d, 10.0 Hz, H-1), 5.95 (d, J 10.0 Hz, H-2), 1.90 (d, J 9.0 Hz, 2H-6), 4.10 (t, J 9.0 Hz, H-7), 5.85 (dd, J 10.3, 2.5 Hz, H-9), 5.80 (ddd, J 10.3, 6.1, 1.7 Hz, H-11), 3.76 (s, H-15), 5.65 (s, H-17), 0.81, 1.10, 1.48 (6H), 1.50 (Me), 7.40 (bs, 2H $\alpha$ -furan), 6.37 (bs H $\beta$ -furan).

#### Methanolic HCl treatment of Carapolide E 4

**Methyl acetal 10.** Carapolide E 4 (50 mg) was dissolved in a mixture of  $\text{CH}_3\text{OH}$  and HCl (8 M) (15 ml) and the solution refluxed for 1h. The reaction mixture was diluted with water and extracted with  $\text{CHCl}_3$ . Column chromatography afforded a single non-crystalline product, the methyl acetal 10 (43 mg). IR ( $\text{CHCl}_3$ )  $\nu_{\text{max}}$  1738, 1720, 1600, 878  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR (200 MHz,  $\text{CDCl}_3$ ) 6.75 (d, J 10 Hz, H-1), 6.08 (d, J 10 Hz, H-2),

4.13 (dd, J 11.0, 6.0 Hz, H-7), 5.85 (dd, J 10.3, 2.5 Hz, H-9), 5.77 (m, H-11), 3.69 (s, H-15), 5.60 (s, H-17), 3.50 (s, OMe), 0.75, 1.12, 1.40, 1.43, 1.58 (Me), 7.35 (m, H-21), 6.35 (m, H-22), 7.35 (m, H-23).

*Carapolide F 5*. Mp 252-255° (ex CHCl<sub>3</sub>/MeOH); [ $\alpha$ ]<sub>D</sub><sup>20</sup> + 105° (c, 3.0 in MeOH) and +128° (c, 0.4 in CHCl<sub>3</sub>); IR (KBr)  $\nu_{\max}$  3490, 3400, 3140, 1760, 1750, 1700, 1650, 1610, 1230, 1070, 1010, 875, 830, 800 cm<sup>-1</sup>. MS *m/z* (%) 530 (M<sup>+</sup>, 10), 514 (5), 470 (20), 452 (30), 412 (28), 396 (28), 376 (100), 365 (80), 343 (60), 329 (75), 303 (50). Found: C, 63.29; H, 6.55. C<sub>28</sub>H<sub>34</sub>O<sub>10</sub> requires C, 63.38; H, 6.46. <sup>1</sup>H and <sup>13</sup>C NMR see Tables 1 and 2.

*Carapolide F Sulphites 11 and 12*. To a solution of carapolide F 5 (10 mg) in dry pyridine (5 drops) was added redistilled SOCl<sub>2</sub> (5 drops). After 15 min, the reaction mixture was warmed to r.t., diluted with water, and extracted with CHCl<sub>3</sub>. Preparative TLC of the crude product afforded two isomeric sulphites 11 (4 mg) and 12 (5 mg). 11 Brown gum: IR (KBr)  $\nu_{\max}$  3142, 1750, 1720, 875 cm<sup>-1</sup>. <sup>1</sup>H NMR (200 MHz, CHCl<sub>3</sub>) 6.71 (d, J 10.0 Hz, H-1), 5.89 (d, J 10.0 Hz, H-2), 2.13 (dd, J 15.5, 3.9 Hz, H-6), 2.59 (dd, J 15.5, 2.0 Hz, H-6 $\beta$ ), 4.58 (dd, J 3.9, 2.0 Hz, H-7), 2.59 (d, J 3.4 Hz, H-9), 5.78 (m, H-11), 1.89 (dd, J 14.6, 5.0 Hz, H-12 $\alpha$ ), 2.38 (dd, J 14.6, 10.3 Hz, H-12 $\beta$ ), 3.59 (s, H-15), 5.72 (s, H-17), 7.45 (m, H-21), 6.32 (m, H-22), 7.45 (m, H-23), 1.22, 1.49, 1.64, 1.67, 2.20 (Me), 2.05 (s, OAc).

The second sulphite 12 was also obtained as a brown gum, IR (KBr)  $\nu_{\max}$  3140, 1752, 1733, 1720, 875 cm<sup>-1</sup>. <sup>1</sup>H NMR (200 MHz, CHCl<sub>3</sub>) 6.73 (d, J 10.0 Hz, H-1), 5.93 (d, J 10 Hz, H-2), 2.21 (dd, J 15.5, 4.0 Hz, H-6 $\alpha$ ), 2.69 (dd, J 15.5, 2.2 Hz, H-6 $\beta$ ), 4.58 (dd, J 4.0, 2.2 Hz, H-7), 2.46 (d, J 3.4 Hz, H-9), 5.49 (m, H-11), 1.90 (dd, J 14.4, 5.0 Hz, H-12 $\alpha$ ), 2.38 (dd, J 14.4, 10.2 Hz, H-12 $\beta$ ), 3.50 (s, H-15), 5.70 (s, H-17), 7.45 (m, H-21), 6.32 (m, H-22), 7.45 (m, H-23), 1.18, 1.55, 1.67 (z), 1.99 (Me), 2.05 (s, OAc).

*Carapolide G 6*. Mp 280° (ex CHCl<sub>3</sub>/MeOH); [ $\alpha$ ]<sub>D</sub><sup>20</sup> + 142° (c, 0.6 in CHCl<sub>3</sub>); IR (KBr)  $\nu_{\max}$  3500, 3140, 1760, 1740, 1715, 1700, 1400, 1080, 880, 845, 805 cm<sup>-1</sup>. MS *m/z* (%) 514 (M<sup>+</sup>, 8), 496 (10), 454 (25), 436 (85), 413 (20), 413 (20), 391 (20), 360 (100), 345 (40), 331 (95), 313 (90), 313 (90), 301 (20). Found: C, 65.28; H, 6.75. C<sub>28</sub>H<sub>34</sub>O<sub>9</sub> requires C, 65.35; H, 6.66. <sup>1</sup>H and <sup>13</sup>C NMR see Tables 2 and 4.

*Carapolide H 7*. Mp 188-190° (ex MeOH); [ $\alpha$ ]<sub>D</sub><sup>20</sup> -58° (c = 0.8 in CHCl<sub>3</sub>); IR (KBr)  $\nu_{\max}$  3140, 1770, 1740, 1720, 1700, 1370, 1225, 1030, 950, 910, 880, 860, 830, 800, 740 cm<sup>-1</sup>. MS *m/z* (%) 543 (M<sup>+</sup> + 1, 3), 542 (M<sup>+</sup>, 9), 482 (6), 422 (5), 364 (6), 351 (5), 190 (15), 174 (12), 161 (10), 108 (42), 43 (100). Found: C, 66.32; H, 7.14. C<sub>30</sub>H<sub>38</sub>O<sub>9</sub> requires C, 66.40; H, 7.06. <sup>1</sup>H and <sup>13</sup>C NMR see Tables 2 and 4.

*Carapolide I 8*. Mp 208-210° (ex EtOAc/hexane); [ $\alpha$ ]<sub>D</sub><sup>20</sup> -46° (c = 1.2 in CHCl<sub>3</sub>). IR  $\nu_{\max}$  1730, 1725, 1370, 1250, 1120, 1025, 875, 805, 780 cm<sup>-1</sup>. MS *m/z* (%) 512 (M<sup>+</sup>, 3), 452 (10), 392 (10), 377 (8), 334 (10), 319 (15), 225 (15), 209 (11), 171 (10), 145 (15), 105 (20), 91 (25), 81 (30), 43 (100). Found: C, 70.38; H, 7.78. C<sub>30</sub>H<sub>40</sub>O<sub>9</sub> requires C, 70.29; H, 7.87. <sup>1</sup>H and <sup>13</sup>C NMR see Tables 2 and 4.

*Evodoulone 9*. Mp 198-200° (lit.<sup>5</sup> 199-200°). IR  $\nu_{\max}$  1745, 1730, 1715, 1680, 1390, 1370, 1240, 1030, 915, 880, 835, 780 cm<sup>-1</sup>.

**Acknowledgements.** We thank Mr. Paul Misili of the Cameroon National Herbarium, Yaoundé, for collection of *C. grandiflora*. Financial support for the work in Yaoundé by the International Programs in the Chemical Sciences (IPICS), Uppsala University, Sweden (Grant No. CAM-02) is gratefully acknowledged.

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(Received in UK 19 January 1994; revised 13 June 1994; accepted 17 June 1994)